

Extensive genetic diversity of Rickettsiales bacteria in multiple mosquito species

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Supplementary Table S1. Bacterial sequences obtained in this study.

Strains	Genes(nt)			Bacteria ^a
	rrs	gltA	groEL	
<i>Anopheles sinensis</i>				
Egg				
WHANSE-2	1400 (99.2%) ^b	- ^c	-	<i>A. platys</i>
WHANSE-2-1	330 (99.1%)	-	-	<i>A. marginale</i>
Larvae				
WHANSL-11-1	343 (100%)	-	-	<i>A. marginale</i>
WHANSL-11	345 (97.0%)	-	-	<i>A. phagocytophilum</i>
WHANSL-24-1	342 (98.5%)	-	-	<i>A. marginale</i>
WHANSL-24	345 (98.6%)	-	-	<i>A. bovis</i>
WHANSL-27-1	1430 (99.6%)	838 (58.5%)	1050 (84.9%)	<i>A. platys</i>
WHANSL-27-2	1380 (99.5%)	-	862 (98.8%)	<i>A. marginale</i>
WHANSL-30	337 (98.8%)	-	-	<i>A. phagocytophilum</i>
WHANSL-34	346 (96.4%)	-	-	<i>A. phagocytophilum</i>
WHANSL-36	353 (98.0%)	-	-	<i>A. phagocytophilum</i>
WHANSL-41	343 (99.1%)	-	-	<i>A. marginale</i>
WHANSL-8-1	342 (99.7%)	-	-	<i>A. marginale</i>
WHANSL-8	1440 (97.8%)	-	-	<i>A. bovis</i>
WHANSL-16	355 (96.1%)	-	-	<i>A. phagocytophilum</i>
WHANSL-LabF50	650 (100%)	-	-	<i>R. japonica</i>
WHANSL-LabF52	650 (100%)	-	-	<i>R. japonica</i>
WHANSL-LabF59	650 (100%)	-	-	<i>R. japonica</i>
Pupae				
WHANSP-48-1	342 (96.8%)	-	-	<i>A. phagocytophilum</i>
WHANSP-48	1402 (99.4%)	-	580 (92.7%)	<i>C. A. boleense</i>
WHANSP-5-1	342 (99.1%)	-	-	<i>A. marginale</i>
WHANSP-60	240 (98.7%)	-	-	<i>C. N. mik</i>
WHANSP-LabF85	1100 (99.6%)	-	-	<i>R. japonica</i>
WHANSP-LabF100	650 (100%)	-	-	<i>R. japonica</i>
WHANSP-LabF101	1100 (100%)	-	-	<i>R. japonica</i>
WHANSP-LabF106	650 (100%)	-	-	<i>R. japonica</i>
Adult				
JXANSA-19-1	400 (99.2)	-	-	<i>A. bovis</i>
JXANSA-19	1434 (99.3%)	832 (57.6%)	1035 (83.6%)	<i>A. platys</i>
JXANSA-2	1378 (99.3%)	-	859 (98.7%)	<i>A. marginale</i>
JXANSA-24	1377 (99.1%)	-	-	<i>A. marginale</i>
JXANSA-30-1	381 (98.4%)	-	-	<i>A. bovis</i>
JXANSA-30	340 (98.8%)	-	-	<i>A. platys</i>
JXANSA-34	333 (99.4%)	-	-	<i>A. marginale</i>
JXANSA-8	333 (100%)	-	-	<i>A. marginale</i>

(Continued)

Strains	Genes(nt)			Bacteria
	<i>rrs</i>	<i>gltA</i>	<i>groEL</i>	
ZJANSA-43	1442 (99.6%)	1186 (99.6%)	862 (98.8%)	<i>A. marginale</i>
WHANSA-12	978 (99.6%)	-	-	<i>A. marginale</i>
WHANSA-13	1428 (99.5%)	-	-	<i>A. marginale</i>
WHANSA-15	1439 (99.9%)	-	-	<i>A. marginale</i>
WHANSA-19	898 (99.8%)	-	-	<i>A. marginale</i>
WHANSA-22	953 (99.6%)	-	-	<i>A. marginale</i>
WHANSA-24-1	1348 (98.5%)	-	625 (86.4%)	<i>A. bovis</i>
WHANSA-24-2	1438 (99.2%)	-	1020 (84.9%)	<i>A. platys</i>
WHANSA-29	1402 (99.7%)	685 (83.2%)	491 (91.0%)	<i>C. A. boleense</i>
WHANSA-29-12	1381 (99.8%)	-	-	<i>A. marginale</i>
WHANSA-36	1349 (99.9%)	-	-	<i>A. marginale</i>
WHANSA-37	1361 (99.9%)	-	-	<i>A. marginale</i>
WHANSA-38	1429 (99.9%)	-	-	<i>A. marginale</i>
WHANSA-39	1432 (99.4%)	-	-	<i>A. marginale</i>
WHANSA-40	1433 (99.2%)	-	-	<i>A. platys</i>
WHANSA-45-1	1393 (98.1%)	-	-	<i>A. platys</i>
WHANSA-45-2	1440 (98.5%)	-	-	<i>A. marginale</i>
WHANSA-47-1	990 (99.8%)	-	-	<i>A. marginale</i>
WHANSA-48	1409 (99.6%)	-	-	<i>A. marginale</i>
WHANSA-52-1	466 (99.8%)	-	-	<i>A. marginale</i>
WHANSA-52	454 (98.4%)	-	-	<i>A. bovis</i>
WHANSA-53-2	526 (98.7%)	-	-	<i>A. marginale</i>
WHANSA-53	525 (99.2%)	-	-	<i>A. bovis</i>
WHANSA-6-1	1436 (98.6%)	-	628 (87.2%)	<i>A. bovis</i>
WHANSA-6-2	1442 (99.7%)	1219 (99.6%)	-	<i>A. marginale</i>
WHANSA-6-3	1437 (99.2%)	-	1050 (84.7%)	<i>A. platys</i>
WHANSA-60-2	595 (99.5%)	-	-	<i>A. marginale</i>
WHANSA-63-1	582 (100%)	-	-	<i>A. marginale</i>
WHANSA-63	519 (99.4%)	-	-	<i>A. platys</i>
WHANSA-7-4	1427 (97.9%)	-	-	<i>A. platys</i>
WHANSA-71-1	516 (99.8%)	-	-	<i>A. marginale</i>
WHANSA-71	555 (99.5%)	-	-	<i>A. platys</i>
WHANSA-79	509 (98.6%)	-	-	<i>A. marginale</i>
WHANSA-8-2	1429 (99.5%)	-	-	<i>A. marginale</i>
WHANSA-8	1430 (99.4%)	-	1020 (84.4%)	<i>A. platys</i>
WHANSA-82-1	573 (98.6%)	-	-	<i>A. bovis</i>
WHANSA-82	658 (99.2%)	-	-	<i>A. marginale</i>
WHANSA-83	398 (99.7%)	-	-	<i>A. marginale</i>
WHANSA-89	395 (99.0%)	-	-	<i>A. marginale</i>

(Continued)

Strains	Genes(nt)			Bacteria
	rrs	gltA	groEL	
WHANSA-92	381 (99.2%)	-	-	<i>A. marginale</i>
WHANSA-122	240 (100%)	-	-	<i>C. A. rodmosense</i>
WHANSA-161	240 (100%)	-	-	<i>C. A. rodmosense</i>
<i>Ehrlichia</i> sp. WHANSA-181	650 (99.8%)	-	-	<i>Ehrlichia</i> sp. EHh317
WHANSA-97	1300 (99.9%)	1000 (97.4%)	400 (96.6%)	<i>R. bellii</i>
WHANSA-146	1300 (98.4%)	-	-	<i>C. R. as</i>
WHANSA-108	240 (98.7%)	-	-	<i>C. N. mik</i>
WHANSA-133	240 (98.7%)	-	-	<i>C. N. mik</i>
WHANSA-LabF24	650 (100%)	-	-	<i>R. japonica</i>
WHANSA-LabP60	650 (99.5%)	-	-	<i>R. japonica</i>
WHANSA-LabP66	650 (100%)	-	-	<i>R. japonica</i>
WHANSA-LabP86	650 (100%)	-	-	<i>R. japonica</i>
WHANSA-LabP95	650 (100%)	-	-	<i>R. japonica</i>
<i>Armigeres subalbatus</i>				
Egg				
WHARSE-LabF5	650 (100%)	-	-	<i>R. japonica</i>
Larvae				
WHARSL-11-1	336 (97.6%)	-	-	<i>A. phagocytophilum</i>
WHARSL-11	343 (97.4%)	-	-	<i>A. bovis</i>
WHARSL-12	342 (97.7%)	-	-	<i>A. phagocytophilum</i>
WHARSL-16-1	374 (98.7%)	-	-	<i>A. phagocytophilum</i>
WHARSL-18-1	361 (99.4%)	-	-	<i>A. marginale</i>
WHARSL-18	340 (97.4%)	-	-	<i>A. phagocytophilum</i>
WHARSL-2-1	389 (99.2%)	-	-	<i>A. phagocytophilum</i>
WHARSL-2	1342 (99.3%)	-	563 (92.0%)	<i>C. A. boleense</i>
WHARSL-21-1	373 (99.2%)	-	-	<i>A. marginale</i>
WHARSL-22	342 (97.4%)	-	-	<i>A. bovis</i>
WHARSL-24	343 (98.0%)	-	-	<i>A. bovis</i>
WHARSL-26-1	351 (99.4%)	-	-	<i>A. marginale</i>
WHARSL-26	336 (97.3%)	-	-	<i>A. phagocytophilum</i>
WHARSL-28-1	372 (98.4%)	-	-	<i>A. bovis</i>
WHARSL-28	970 (99.6%)	-	-	<i>A. marginale</i>
WHARSL-3-1	375 (99.7%)	-	-	<i>A. marginale</i>
WHARSL-3	324 (97.8%)	-	-	<i>A. phagocytophilum</i>
WHARSL-30	1381 (99.3%)	826 (57.6%)	1041 (84.9%)	<i>A. platys</i>
WHARSL-32	333 (98.8%)	-	-	<i>A. platys</i>
WHARSL-33	343 (97.1%)	-	-	<i>A. bovis</i>
WHARSL-34	343 (96.8%)	-	-	<i>A. phagocytophilum</i>
WHARSL-35	332 (97.6%)	-	-	<i>A. phagocytophilum</i>

(Continued)

Strains	Genes(nt)			Bacteria
	rrs	gltA	groEL	
WHARSL-38-1	1376 (99.8%)	1180 (99.5%)	850 (98.6%)	<i>A. marginale</i>
WHARSL-38-2	1441 (98.5%)	-	628 (86.9%)	<i>A. bovis</i>
WHARSL-9	335 (97.9%)	-	-	<i>A. phagocytophilum</i>
WHARSL-LabF8	650 (100%)	-	-	<i>R. japonica</i>
WHARSL-LabF12	650 (100%)	-	-	<i>R. japonica</i>
WHARSL-LabF14	650 (100%)	-	-	<i>R. japonica</i>
WHARSL-LabF21	650 (100%)	-	-	<i>R. japonica</i>
WHARSL-LabF34	650 (100%)	-	-	<i>R. japonica</i>
WHARSL-LabF56	650 (100%)	-	-	<i>R. japonica</i>
WHARSL-LabF115	650 (100%)	-	-	<i>R. japonica</i>
WHARSL-LabF130	650 (100%)	-	-	<i>R. japonica</i>
Pupae				
WHARSP-17	1381 (99.4%)	-	993 (85.1%)	<i>A. platys</i>
WHARSP-19	1378 (99.9%)	-	850 (98.6%)	<i>A. marginale</i>
WHARSP-2-1	371 (98.9%)	-	-	<i>A. bovis</i>
WHARSP-30-1	372 (98.7%)	-	-	<i>A. bovis</i>
WHARSP-30-2	343 (98.8%)	-	-	<i>A. marginale</i>
WHARSP-38	1372 (99.6%)	954 (84.2%)	-	<i>C. A. boleense</i>
WHARSP-LabF3	650 (100%)	-	-	<i>R. japonica</i>
WHARSP-LabF11	650 (100%)	-	-	<i>R. japonica</i>
WHARSP-LabF18	650 (100%)	-	-	<i>R. japonica</i>
WHARSP-LabF21	650 (100%)	-	-	<i>R. japonica</i>
WHARSP-LabF23	650 (100%)	-	-	<i>R. japonica</i>
WHARSP-LabF39	650 (100%)	-	-	<i>R. japonica</i>
WHARSP-LabF42	650 (100%)	-	-	<i>R. japonica</i>
Adult				
JXARSA-1	348 (99.4%)	-	-	<i>A. phagocytophilum</i>
JXARSA-15	354 (99.2%)	-	-	<i>A. phagocytophilum</i>
JXARSA-2	356 (99.1%)	-	-	<i>A. phagocytophilum</i>
JXARSA-29	1359 (99.2%)	832 (58.2%)	866 (86.7%)	<i>A. platys</i>
JXARSA-32	343 (99.4%)	-	-	<i>A. phagocytophilum</i>
JXARSA-33	1360 (99.3%)	679 (83.2%)	530 (92.6%)	<i>C. A. boleense</i>
JXARSA-5	1371 (99.1%)	678 (83.2%)	-	<i>C. A. boleense</i>
WHARSA-14-1	321 (99.4%)	-	-	<i>A. marginale</i>
WHARSA-14	1002 (98.8%)	-	1017 (84.2%)	<i>A. platys</i>
WHARSA-30-1	361 (98.9%)	-	-	<i>A. bovis</i>
WHARSA-30	1380 (99.9%)	-	850 (98.0%)	<i>A. marginale</i>
WHARSA-40-1	1380 (99.7%)	-	835 (98.6%)	<i>A. marginale</i>
WHARSA-40-2	1349 (98.5%)	-	628 (86.3%)	<i>A. bovis</i>

(Continued)

Strains	Genes(nt)			Bacteria
	rrs	gltA	groEL	
WHARSA-47-1	1380 (99.5%)	-	-	<i>A. platys</i>
WHARSA-47-2	1378 (99.9%)	-	-	<i>A. marginale</i>
WHARSA-5	366 (98.9%)	-	-	<i>A. phagocytophilum</i>
WHARSA-7	1380 (99.6%)	-	1047 (83.7%)	<i>A. platys</i>
ZJARSA-11	358 (98.9%)	-	-	<i>A. platys</i>
ZJARSA-2	349 (98.6%)	-	-	<i>A. phagocytophilum</i>
ZJARSA-4	1393 (99.3%)	685 (82.5%)	491 (91.0%)	<i>C. A. boleense</i>
ZJARSA-8	1380 (99.6)	618 (56.5%)	987 (84.6%)	<i>A. platys</i>
WHARSA-80	240 (100%)	-	-	<i>C. A. rodmosense</i>
WHARSA-86	650 (99.1%)	-	-	<i>A. phagocytophilum</i>
WHARSA-103	650 (99.2%)	-	-	<i>A. phagocytophilum</i>
WHARSA-106	650 (98.9%)	-	-	<i>A. bovis</i>
WHARSA-121	650 (99.2%)	-	-	<i>A. bovis</i>
WHARSA-132	650 (99.2%)	-	-	<i>A. bovis</i>
WHARSA-87-1	650 (99.3%)	-	-	<i>A. phagocytophilum</i>
<i>Ehrlichia</i> sp. WHARSA-87	650 (99.6%)	-	450 (89.4%)	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHARSA-128	650 (99.4%)	-	-	<i>Ehrlichia</i> sp. NS101
<i>Ehrlichia</i> sp. WHARSA-134	1110 (99.7%)	-	-	<i>Ehrlichia</i> sp. EHh317
WHARSA-85	1110 (99.3%)	-	-	<i>C. N. mik</i>
WHARSA-107	650 (99.6%)	-	-	<i>C. N. mik</i>
WHARSA-115	650 (99.6%)	-	-	<i>C. N. mik</i>
WHARSA-118	650 (99.6%)	-	-	<i>C. N. mik</i>
WHARSA-135	650 (99.6%)	-	-	<i>C. N. mik</i>
WHARSA-140	650 (99.6%)	-	-	<i>C. N. mik</i>
WHARSA-142	650 (99.6%)	-	-	<i>C. N. mik</i>
WHARSA-LabF12	650 (100%)	-	-	<i>R. japonica</i>
WHARSA-LabP24	650 (100%)	-	-	<i>R. japonica</i>
WHARSA-LabP51	650 (100%)	-	-	<i>R. japonica</i>
WHARSA-LabP70	650 (100%)	-	-	<i>R. japonica</i>
WHARSA-LabP71	650 (100%)	-	-	<i>R. japonica</i>
<i>Aedes albopictus</i>				
Larvae				
WHAEAL-10	357 (99.2%)	-	-	<i>A. phagocytophilum</i>
WHAEAL-14	346 (99.3%)	-	-	<i>A. phagocytophilum</i>
WHAEAL-16	357 (99.3%)	-	-	<i>A. phagocytophilum</i>
WHAEAL-17-1	1346 (99.3%)	1201 (99.5%)	856 (98.6%)	<i>A. marginale</i>
WHAEAL-17-2	938 (99.2%)	832 (57.9%)	1014 (84.5%)	<i>A. platys</i>
WHAEAL-23	356 (99.2%)	-	-	<i>A. phagocytophilum</i>
WHAEAL-25	339 (99.4%)	-	-	<i>A. phagocytophilum</i>

(Continued)

Strains	Genes(nt)			Bacteria
	rrs	gltA	groEL	
WHAEAL-45	1356 (99.2%)	685 (82.5%)	-	<i>C. A. boleense</i>
WHAEAL-7-1	345 (99.4%)	-	-	<i>A. bovis</i>
WHAEAL-7-2	374 (99.2%)	-	-	<i>A. marginale</i>
WHAEAL-69	650 (98.8%)	-	-	<i>A. phagocytophilum</i>
WHAEAL-81	650 (99.8%)	-	-	<i>A. marginale</i>
WHAEAL-84	550 (99.1%)	-	-	<i>A. bovis</i>
WHAEAL-109	650 (99.1%)	-	-	<i>A. bovis</i>
WHAEAL-112	240 (100%)	-	-	<i>C. A. rodmosense</i>
WHAEAL-56	535 (99.6%)	-	-	<i>A. marginale</i>
WHAEAL-79-1	668 (99.6%)	-	-	<i>A. phagocytophilum</i>
WHAEAL-79-2	667 (98.9%)	-	-	<i>A. bovis</i>
<i>Ehrlichia</i> sp. WHAEAL-56-1	650 (99.9%)	-	-	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHAEAL-79	650 (100%)	-	450 (89.4%)	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHAEAL-110	650 (99.9%)	-	-	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHAEAL-113	650 (99.2%)	-	-	<i>Ehrlichia</i> sp. NS101
<i>Ehrlichia</i> sp. WHAEAL-132	650 (99.1%)	-	450 (89.2%)	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHAEAL-139	1110 (99.2%)	-	-	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHAEAL-144	1110 (99.3%)	-	-	<i>Ehrlichia</i> sp. EHh317
WHAEAL-125	1110 (98.8%)	-	-	<i>C. N. mik</i>
WHAEAL-130	650 (99.6%)	-	-	<i>C. N. mik</i>
WHAEAL-LabF6	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF18	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF21	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF34	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF37	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF39	1100 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF49	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF50	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF52	650 (100%)	-	-	<i>R. japonica</i>
WHAEAL-LabF54	650 (100%)	-	-	<i>R. japonica</i>
<i>Ehrlichia</i> sp. WHAEAL-LabF19	650 (99.4%)	-	-	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHAEAL-LabF20	650 (99.4%)	-	-	<i>Ehrlichia</i> sp. EHh317
Pupae				
WHAEAP-26	938 (98.9%)	823 (57.7%)	1047 (83.5%)	<i>A. platys</i>
WHAEAP-33	1380 (99.7%)	-	850 (98.4%)	<i>A. marginale</i>
WHAEAP-67	240 (100%)	-	-	<i>C. A. rodmosense</i>
<i>Ehrlichia</i> sp. WHAEAP-65	240 (100%)	-	-	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHAEAP-69	240 (100%)	-	-	<i>Ehrlichia</i> sp. EHh317
WHAEAP-55	240 (98.7%)	-	-	<i>C. N. mik</i>

(Continued)

Strains	Genes(nt)			Bacteria
	rrs	gltA	groEL	
WHAEAP-61	240 (98.7%)	-	-	<i>C. N. mik</i>
WHAEAP-LabF43	650 (100%)	-	-	<i>R. japonica</i>
WHAEAP-LabF45	650 (100%)	-	-	<i>R. japonica</i>
Adult				
ZJAEAA-1	1382 (99.7%)	1195 (99.5%)	817 (98.5%)	<i>A. marginale</i>
ZJAEAA-4	345 (99.4%)	-	-	<i>A. phagocytophilum</i>
ZJAEAA-5-1	377 (99.5%)	-	-	<i>A. marginale</i>
ZJAEAA-5	1379 (99.4%)	666 (84.5%)	491 (92.0%)	<i>C. A. boleense</i>
WHAEAA-39	240 (98.7%)	-	-	<i>C. N. mik</i>
WHAEAA-LabF26	1100 (99.8%)	-	-	<i>R. japonica</i>
WHAEAA-LabP5	650 (100%)	-	-	<i>R. japonica</i>
WHAEAA-LabP12	650 (100%)	-	-	<i>R. japonica</i>
<i>Ehrlichia</i> sp. WHAEAA-LabP15	650 (99.6%)	-	-	<i>Ehrlichia</i> sp. EHh317
WHAEAA-LabP33	650 (100%)	-	-	<i>R. japonica</i>
WHAEAA-LabP44	650 (99.5%)	-	-	<i>R. japonica</i>
WHAEAA-LabP57	650 (99.5%)	-	-	<i>R. japonica</i>
WHAEAA-LabP58	650 (99.5%)	-	-	<i>R. japonica</i>
WHAEAA-LabP68	650 (100%)	-	-	<i>R. japonica</i>
WHAEAA-LabP82	650 (99.5%)	-	-	<i>R. japonica</i>
<i>Culex quinquefasciatus</i>				
Egg				
WHCUQE-LabF3	650 (100%)	-	-	<i>R. japonica</i>
WHCUQE-LabF19	650 (100%)	-	-	<i>R. japonica</i>
Larvae				
WHCUQL-18	356 (97.2%)	-	-	<i>A. phagocytophilum</i>
WHCUQL-2	356 (98.9%)	-	-	<i>A. phagocytophilum</i>
WHCUQL-4	356 (98.0%)	-	-	<i>A. phagocytophilum</i>
WHCUQL-46	1323 (99.4%)	682 (82.5%)	-	<i>C. A. boleense</i>
WHCUQL-65	240 (100%)	-	-	<i>C. A. rodmosense</i>
WHCUQL-98	240 (100%)	-	-	<i>A. marginale</i>
WHCUQL-112	240 (97.3%)	-	-	<i>C. A. rodmosense</i>
WHCUQL-118	240 (98.3%)	-	-	<i>A. marginale</i>
WHCUQL-83	240 (100%)	-	-	<i>E. chaffeensis</i>
WHCUQL-53	240 (98.7%)	-	-	<i>C. N. mik</i>
WHCUQL-110	240 (98.7%)	-	-	<i>C. N. mik</i>
WHCUQL-LabF8	650 (100%)	-	-	<i>R. japonica</i>
WHCUQL-LabF14	650 (100%)	-	-	<i>R. japonica</i>
WHCUQL-LabF89	650 (100%)	-	-	<i>R. japonica</i>
WHCUQL-LabP6	650 (100%)	-	-	<i>R. japonica</i>

(Continued)

Strains	Genes(nt)			Bacteria
	rrs	gltA	groEL	
WHCUQL-LabP11	650 (99.5%)	-	-	<i>R. japonica</i>
WHCUQL-LabP27	650 (100%)	-	-	<i>R. japonica</i>
WHCUQL-LabP34	650 (99.5%)	-	-	<i>R. japonica</i>
WHCUQL-LabP39	650 (100%)	-	-	<i>R. japonica</i>
WHCUQL-LabP60	650 (100%)	-	-	<i>R. japonica</i>
Pupae				
WHCUQP-88	650 (99.2%)	-	-	<i>A. bovis</i>
<i>Ehrlichia</i> sp. WHCUQP-70	240 (100%)	-	-	<i>Ehrlichia</i> sp. EHh317
WHCUQP-78	240 (98.7%)	-	-	<i>C. N. mik</i>
WHCUQP-LabF5	650 (100%)	-	-	<i>R. japonica</i>
WHCUQP-LabF7	650 (100%)	-	-	<i>R. japonica</i>
WHCUQP-LabF68	1100 (99.5%)	-	-	<i>R. japonica</i>
WHCUQP-LabF78	650 (100%)	-	-	<i>R. japonica</i>
WHCUQP-LabF79	650 (100%)	-	-	<i>R. japonica</i>
WHCUQP-LabF81	650 (100%)	-	-	<i>R. japonica</i>
Adult				
WHCUQA-73	240 (100%)	-	-	<i>C. A. rodmosense</i>
WHCUQA-81	240 (100%)	-	-	<i>C. A. rodmosense</i>
WHCUQA-68	240 (100%)	-	-	<i>E. chaffeensis</i>
<i>Ehrlichia</i> sp. WHCUQA-72	240 (100%)	-	-	<i>Ehrlichia</i> sp. EHh317
<i>Ehrlichia</i> sp. WHCUQA-111	240 (100%)	-	-	<i>Ehrlichia</i> sp. EHh317
WHCUQA-97	1300 (99.8%)	1000 (100%)	700 (99.6%)	<i>R. monacensis</i>
WHCUQA-63	240 (98.7%)	-	-	<i>C. N. mik</i>
WHCUQA-LabF56	650 (100%)	-	-	<i>R. japonica</i>
WHCUQA-LabP13	650 (99.5%)	-	-	<i>R. japonica</i>
WHCUQA-LabP36	650 (99.5%)	-	-	<i>R. japonica</i>
WHCUQA-LabP43	650 (99.5%)	-	-	<i>R. japonica</i>
WHCUQA-LabP48	650 (99.5%)	-	-	<i>R. japonica</i>
WHCUQA-LabP88	650 (100%)	-	-	<i>R. japonica</i>
<i>Culex tritaeniorhynchus</i>				
Egg				
WHCUTE-LabF3	650 (100%)	-	-	<i>R. sibirica</i>
WHCUTE-LabF5	650 (99.8%)	-	-	<i>R. monacensis</i>
Larvae				
WHCUTL-97	650 (99.2%)	-	-	<i>A. platys</i>
WHCUTL-131	650 (98.3%)	-	-	<i>A. platys</i>
WHCUTL-65	650 (97.6%)	-	-	<i>C. R. ct</i>
WHCUTL-121	240 (98.7%)	-	-	<i>C. N. mik</i>
WHCUTL-LabF11	650 (100%)	-	-	<i>R. sibirica</i>

(Continued)

Strains	Genes(nt)			Bacteria
	rrs	gltA	groEL	
WHCUTL-LabF24	650 (99.8%)	-	-	<i>R. monacensis</i>
WHCUTL-LabF28	1100 (100%)	-	-	<i>R. sibirica</i>
WHCUTL-LabF41	650 (100%)	-	-	<i>R. japonica</i>
WHCUTL-LabF43	650 (100%)	-	-	<i>R. japonica</i>
WHCUTL-LabF80	1100 (99.9%)	-	-	<i>R. monacensis</i>
Pupae				
WHCUTP-70	240 (100%)	-	-	<i>E. chaffeensis</i>
WHCUTP-78	240 (98.7%)	-	-	<i>C. N. mik</i>
WHCUTP-122	240 (98.7%)	-	-	<i>C. N. mik</i>
WHCUTP-LabF7	650 (100%)	-	-	<i>R. sibirica</i>
WHCUTP-LabF9	1100 (99.8%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF18	1100 (99.9%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF23	650 (99.8%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF33	1100 (100%)	-	-	<i>R. japonica</i>
WHCUTP-LabF61	1100 (99.8%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF64	650 (99.8%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF94	1100 (99.8%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF97	1100 (99.9%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF110	1100 (99.8%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF112	1100 (99.9%)	-	-	<i>R. monacensis</i>
WHCUTP-LabF118	650 (99.8%)	-	-	<i>R. monacensis</i>
Adult				
WHCUTA-25	1381 (99.6%)	-	-	<i>A. marginale</i>
WHCUTA-8	1381 (99.4%)	-	-	<i>A. marginale</i>
ZJCUTA-10	1370 (99.6%)	-	-	<i>A. marginale</i>
ZJCUTA-2	1379 (99.3%)	-	-	<i>A. marginale</i>
ZJCUTA-30	1351 (99.8%)	-	-	<i>A. marginale</i>
ZJCUTA-5	1380 (99.6%)	-	-	<i>A. marginale</i>
ZJCUTA-6	1380 (99.7%)	-	-	<i>A. marginale</i>
WHCUTA-99	650 (99.1%)	-	-	<i>A. phagocytophilum</i>
WHCUTA-111	650 (99.2%)	-	-	<i>A. platys</i>
WHCUTA-122	650 (99.4%)	-	-	<i>A. platys</i>
WHCUTA-123	650 (98.6%)	-	-	<i>A. platys</i>
WHCUTA-137	650 (99.4%)	-	-	<i>A. platys</i>
WHCUTA-141	650 (98.5%)	-	-	<i>A. platys</i>
WHCUTA-121	1300 (99.3%)	1000 (76.7%)	-	<i>R. nc</i>
WHCUTA-130	1300 (99.3%)	1000 (76.7%)	-	<i>R. nc</i>
WHCUTA-LabF5	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF11	1100 (100%)	-	-	<i>R. japonica</i>

(Continued)

Strains	Genes(nt)			Bacteria
	<i>rrs</i>	<i>gltA</i>	<i>groEL</i>	
WHCUTA-LabF12	1100 (99.9%)	-	-	<i>R. japonica</i>
WHCUTA-LabF13	650 (99.7%)	-	-	<i>R. sibirica</i>
WHCUTA-LabF15	1100 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF20	1100 (99.7%)	-	-	<i>R. monacensis</i>
WHCUTA-LabF33	1100 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF34	1100 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF35	650 (99.7%)	-	-	<i>R. sibirica</i>
WHCUTA-LabF42	1100 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF46	650 (99.7%)	-	-	<i>R. sibirica</i>
WHCUTA-LabF47	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF51	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF53	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF54	1100 (99.9%)	-	-	<i>R. japonica</i>
WHCUTA-LabF59	650 (99.7%)	-	-	<i>R. sibirica</i>
WHCUTA-LabF63	1100 (99.8%)	-	-	<i>R. japonica</i>
WHCUTA-LabF64	1100 (99.8%)	-	-	<i>R. sibirica</i>
WHCUTA-LabF70	1100 (99.7%)	-	-	<i>R. sibirica</i>
WHCUTA-LabF71	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF84	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabF91	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabP31	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabP52	650 (100%)	-	-	<i>R. japonica</i>
WHCUTA-LabP114	650 (99.6%)	-	-	<i>R. japonica</i>
WHCUTA-LabP157	650 (99.5%)	-	-	<i>R. monacensis</i>
WHCUTA-LabP162	650 (100%)	-	-	<i>R. sibirica</i>

^aThe abbreviation is the same as in Tables 1 and 3.

^bNucleotide sequence identity compared to the reference sequences from GenBank.

^c“-”, not available.

Supplementary Table S2. Prevalence of Rickettsiales in adult mosquito sampled from three regions of China.

Species	Location	Years	<i>Anaplasma</i> ^a						<i>Ehrlichia</i>				<i>Rickettsia</i>			<i>C.N.mik</i>	Total
			<i>A.bov</i>	<i>C.A.bol</i>	<i>A.mar</i>	<i>A.pha</i>	<i>A.pla</i>	<i>C.A.rod</i>	<i>E.cha</i>	<i>E.ehh</i>	<i>E.ns</i>	<i>C.R.as</i>	<i>R.bel</i>	<i>R.mon</i>	<i>R.nc</i>		
Ae.a.	Hubei	2015	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1/144
	Zhejiang	2014	0	1	2	1	0	0	0	0	0	0	0	0	0	0	4/6
An.s.	Hubei	2014	5	1	25	0	8	0	0	0	0	0	0	0	0	0	39/96
		2015	0	0	0	0	0	2	0	1	0	1	1	0	0	2	7/96
Jiangxi	Jiangxi	2014	2	0	4	0	2	0	0	0	0	0	0	0	0	0	8/38
	Zhejiang	2014	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1/48
Ar.s.	Hubei	2014	2	0	4	1	3	0	0	0	0	0	0	0	0	0	10/48
		2015	3	0	0	3	0	1	0	2	1	0	0	0	0	7	17/55
Cu.q.	Jiangxi	2014	0	2	0	4	1	0	0	0	0	0	0	0	0	0	7/42
	Zhejiang	2014	0	1	0	1	2	0	0	0	0	0	0	0	0	0	4/14
Cu.t.	Hubei	2014	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0/48
		2015	0	0	0	0	0	2	1	2	0	0	0	1	0	1	7/96
Total			12	5	43	11	21	5	1	5	1	1	1	1	2	11	120/971

^a Abbreviations are the same as those in Table 1.

Supplementary Table S3. Prevalence of Rickettsiales in eggs, larvae and pupae sampled from Hubei Province, China.

Species	Years	Anaplasma ^a					Ehrlichia		Rickettsia		C.N.mik	Total	
		A.bov	C.A.bol	A.pla	A.mar	A.pha	C.A.rod	E.cha	E.ehh	E.ns	C.R.ct		
Egg													
<i>Ae.a</i>	2014	0	0	0	0	0	0	0	0	0	0	0	0/0
	2015	0	0	0	0	0	0	0	0	0	0	0	0/0
<i>An.s</i>	2014	0	0	0	1	1	0	0	0	0	0	0	2/4
	2015	0	0	0	0	0	0	0	0	0	0	0	0/5
<i>Ar.s</i>	2014	0	0	0	0	0	0	0	0	0	0	0	0/0
	2015	0	0	0	0	0	0	0	0	0	0	0	0/0
<i>Cu.q</i>	2014	0	0	0	0	0	0	0	0	0	0	0	0/0
	2015	0	0	0	0	0	0	0	0	0	0	0	0/0
<i>Cu.t</i>	2014	0	0	0	0	0	0	0	0	0	0	0	0/10
	2015	0	0	0	0	0	0	0	0	0	0	0	0/0
Subtotal		0	0	0	1	1	0	0	0	0	0	0	2/19
Larvae													
<i>Ae.a</i>	2014	1	1	1	2	5	0	0	0	0	0	0	10/48
	2015	3	0	0	2	2	1	0	6	1	0	2	17/96
<i>An.s</i>	2014	2	0	1	5	5	0	0	0	0	0	0	13/48
	2015	0	0	0	0	0	0	0	0	0	0	0	0/40
<i>Ar.s</i>	2014	6	1	2	6	10	0	0	0	0	0	0	25/48
	2015	0	1	0	0	3	0	0	0	0	0	0	4/48
<i>Cu.q</i>	2014	0	1	0	0	2	0	2	1	0	0	0	2
	2015	0	0	0	2	0	0	0	0	0	0	0	7/96
<i>Cu.t</i>	2014	0	0	0	0	0	0	0	0	0	0	0	0/96
	2015	0	0	2	0	0	0	0	0	0	1	1	4/48
Subtotal		12	3	6	17	25	3	1	6	1	1	5	80/528
Pupae													
<i>Ae.a</i>	2014	0	0	1	1	0	0	0	0	0	0	0	2/48
	2015	0	0	0	0	0	1	0	2	0	0	2	5/96
<i>An.s</i>	2014	0	1	1	1	0	0	0	0	0	0	0	3/48
	2015	0	0	0	0	0	0	0	0	0	0	1	1/40
<i>Ar.s</i>	2014	2	1	1	2	0	0	0	0	0	0	0	6/48
	2015	1	0	0	0	0	0	0	1	0	0	1	3/144
<i>Cu.t</i>	2014	0	0	0	0	0	0	0	0	0	0	0	0/48
	2015	0	0	0	0	0	0	1	0	0	0	2	3/82
Subtotal		3	2	3	4	0	1	1	3	0	0	6	23/554

^a Abbreviations are the same as those in Tables 1 and 3.

Supplementary Table S4. Primers used in this study.

Bacteria	gene	Sequences (5'→3')	Position ^a	Reference
Rickettsiales	<i>rrs</i>	Eh-out1: TTGAGAGAGTTGATCCTGGCTCAGAACG(+)	1-27	26
		Eh-out2: CACCTCTACACTAGGAATTCCGCTATC(-)	653-627	26
		Eh-gs1: GTAATAACTGTATAATCCCTG(+)	167-187	26
		Eh-gs2: GTACCGTCATTATCTTCCCTA(-)	448-428	26
<i>Anaplasma</i>	<i>rrs</i>	GTTTGATCCTGGCTCAGAA (+)	15-33	This study
		TACACTAGGAATTCCGCTATC (-)	652-632	This study
		CTAATACCGTATATGCYC (+)	173-190	This study
		GTACCGTCATTATCWTCC (-)	453-436	This study
<i>Anaplasma</i>	<i>rrs</i>	GGATAGCCACTRGAARTGGTG(+)	151-171	This study
		CGTGCTGACTTGACATCAT(-)	1172-1154	This study
		CATCTCACGACACGAGCTG(-)	1049-1031	This study
		CTGCTCTGGTCCGGTACTGAC(+)	700-719	This study
		TGGTCCGGTACTGACRCT(+)	705-722	This study
		TGCCTCCTTDCGGTTGGC(-)	1425-1408	This study
<i>Ehrlichia</i>	<i>rrs</i>	GAATAGCCATTAGAAATGATG(+)	155-175	This study
		GTCAGTATCGAACCAAGATAG(-)	723-704	This study
		GTATCGAACCAAGATAGCCG(-)	719-701	This study
		CGGCTATCTGGTTCGATAC(+)	701-719	This study
		CTATCTGGTTCGATACTGAC(+)	704-723	This study
		GCTTCCCTKCGGTTAGCAC(-)	1427-1409	This study
<i>Rickettsia</i>	<i>rrs</i>	GTACCGAATAACTTTAGAAAT(+)	147-168	This study
		CATGATGACTTGACRTCGT(-)	1175-1157	This study
		CATCTCACGACACGAGCTG(-)	1052-1034	This study
		GAAGGCGRTCAYTRGGCT(+)	693-711	This study
		GRTCAYTRGGCTRCAACTG(+)	699-718	This study
		CTGCCCTTGCCTAGCT(-)	1427-1410	This study
<i>Ca. Neoehrlichia</i>	<i>rrs</i>	GACAGGTAATACCRTATAATCCCT(+)	123-146	This study
		GTCAGAACTGAGCCAGATAG(-)	675-656	This study
		CTGAGCCAGATAGTCGC(-)	668-652	This study
		GCGACTATCTGGCTCAG(+)	652-668	This study
		CTATCTGGCTCAGTTCTGAC(+)	656-675	This study
		TGCCTCCTACGGTTAG(-)	1382-1366	This study
<i>A. platys</i>	<i>groEL</i>	AGTCGATTAGGGAAAGTAGTAC(+)	38-58	This study
		AGGATGGCTACAAGGTAATG(+)	149-168	This study
		GCGTCCTCTACTCTGTCTT(-)	1199-1181	This study
<i>A. marginale</i>	<i>groEL</i>	ACATGCTCCATACTGACTGC(+)	268-287	This study
		AGATGAGATTGCACAGGTTG(+)	420-439	This study
		AGATGCAAGCGTGTATAGCAG(-)	1281-1261	This study
<i>A. bovis</i>	<i>groEL</i>	GCTGCTGCGATAGCTAATAT(+)	196-215	This study
		GATGGTACCACTACTTGTCA(+)	256-276	This study
		CTATGACTGCTATGTCACCA(-)	883-864	This study

(Continued)

Bacteria	Genes	Sequences (5'→3')	Position	Reference
<i>Ca. A. boleense</i>	<i>groEL</i>	TAGAAGACGCGGTAGGCT(+)	47-64	This study
		GTACTGCAGGCCCTAAAG(+)	65-82	This study
		AACGTTCTCCAATATGGGAAG(-)	679-699	This study
<i>Ehrlichia</i> sp. EHh317	<i>groEL</i>	GAAGATGCTGTAGGRTGTACDGC(+)	67-89	This study
		ATTRCTCARAGTGCTTCHCARTG(+)	217-239	This study
		AGHGCTTCWCCTTCYACATCYTC(-)	776-754	This study
<i>Rickettsia</i>	<i>groEL</i>	CCATTACATGATAGAATTGCAAT(+)	<i>groES</i>	This study
		GAATTGCAATAAAGCCTATCG(+)	<i>groES</i>	This study
		CCATCATTGCTTTCTTCTATC(-)	868-847	This study
<i>A. platys</i>	<i>gltA</i>	TGRAAGAAAAGCTGTTTG(+)	2-21	This study
		AGCTRTTTTRGAGTGYGGAG(+)	12-31	This study
		GCTCTRGGRTCATARCTYTT(-)	926-907	This study
<i>A. mraginale</i>	<i>gltA</i>	TGGTAGAAAAAGCGATTTAG(+)	2-22	This study
		ATAAGCTTGCCCGTTATGC(+)	40-48	This study
		CCGGTATAAAGTTGGCGT(-)	1235-1218	This study
<i>Ca. A. boleense</i>	<i>gltA</i>	ATGTCTACTGCRGCWTGC(+)	1-18	This study
		TACTGCRGCTWGCAGGTCT (+)	6-24	This study
		CAGYAGTTCTMGCTAATG(-)	1012-996	This study
<i>Rickettsia</i>	<i>gltA</i>	CCGGGYTTTATGTCTACTGC(+)	151-170	This study
		CTTTATGTCTACTGCKTCTTG(+)	156-176	This study
		AGCTGTCTWGGTCTGCTGATT(-)	1259-1239	This study

^aThe numbers correspond to genome positions of reference sequences from GenBank.



Supplementary Figure S1. Map showing the location of the collection areas in Hubei, Jiangxi, and Zhejiang provinces, China. The areas marked with blue represented the locations of Wuhan, Cixi and Yudu, respectively. The figure was generated using ArcGIS (version 10.0; <http://www.arcgis.com/feature/>).